

## 14-120ACL: RIG-I/NF-κB Lleeporter™ Luciferase Reporter-HEK293T Cell Line

**Application :** Functional Assay

### Description

The RIG-I Lleeporter™ Luciferase reporter cell line is a stably transfected HEK 293T cell line which expresses human retinoic acid-induced protein-I (RIG-I) and Renilla luciferase reporter gene under the transcriptional control of the NF-κB response element. As a dsRNA helicase enzyme, RIG-I is encoded by the DDX58 gene. RIG-I is one of the RIG-I-like receptors (RLRs) that are a family of DExD/H box RNA helicases including RIG-I, MDA5 and LPG2, which play a major role in pathogen sensing of RNA virus infection to initiate and modulate antiviral immunity. RLR expression is typically maintained at low levels in resting cells but is greatly increased during inflammation, specifically with IFN exposure and after virus infection. RIG-I detects cytoplasmic dsRNA generated during viral replication unlike Toll-like receptor 3 (TLR3) which can detect phagocytosed dsRNA in endosomes. RIG-I also responds to poly(I:C), the synthetic analog of viral dsRNA. The RIG-I activation by poly(I:C) is shown in Figure 1.

### Product Info

<b>Amount :</b>	1 Vial
<b>Content :</b>	Each vial contains $2 \sim 3 \times 10^6$ cells in 1 ml of 90% FBS + 10% DMSO.
<b>Storage condition :</b>	Immediately upon receipt, store in liquid nitrogen.

### Application Note

#### Application:

- Monitor the RIG-I signaling pathway activity.
- Screen for activators or inhibitors of the RIG-I signaling pathway.

#### Culture conditions:

Cells should be grown at 37°C with 5% CO<sub>2</sub> using DMEM medium (w/ L-Glutamine, 4.5g/L Glucose and Sodium Pyruvate) supplemented with 10% heat-inactivated FBS and 1% Pen/Strep, plus 2 µg/ml of Puromycin and 5 µg/ml Blasticidin (Note: Puromycin and Blasticidin can be omitted during the reporter cell assays).

It is recommended to quickly thaw the frozen cells upon receipt or from liquid nitrogen in a 37°C water-bath, transfer to a tube containing 10 ml of growth medium without Puromycin and Blasticidin, spin down cells, resuspend cells in pre-warmed growth medium without Puromycin and Blasticidin, transfer resuspended cells to T25 flask and culture in 37°C-CO<sub>2</sub> incubator.

Leave the T25 flask in the incubator for 1~3 days without disturbing or changing the medium until cells completely recover viability and become adherent. Once cells are over 90% adherent, remove growth medium and passage the cells through trypsinization and centrifugation. At first passage, switch to growth medium containing Puromycin and Blasticidin. Cells should be split before they reach complete confluence.

To passage the cells, detach cells from culture vessel with Trypsin/EDTA, add complete growth medium and transfer to a tube, spin down cells, resuspend cells and seed appropriate aliquots of cells suspension into new culture vessels. Subcultivation ration = 1:10 to 1:20 weekly. To achieve satisfactory results, cells should not be passaged over 16 times.

## Functional validation:

### A. Response of RIG-I Leepor<sup>TM</sup> - HEK293T cells to Poly(I:C).

1. Harvest RIG-I Leepor<sup>TM</sup> - HEK293T cells and seed cells into a white solid-bottom 96-well microplate in 100  $\mu$ l of growth medium at  $5 \times 10^4$  cells/well.
2. Incubate cells at 37°C in a CO<sub>2</sub> incubator for overnight.
3. The next day, stimulate cells with various concentrations of the poly(I:C) packed with lipofectamine 2000, which was prepared as follows:
  - i) A 50  $\mu$ l of 10 mg/ml poly(I:C) (= 500  $\mu$ g total) is preincubated in 50  $\mu$ l Opti-MEM (Life Technologies) for 5 min.
  - ii) Similarly, a 20  $\mu$ l Lipofectamine 2000 (Life Technologies) is preincubated in 80  $\mu$ l Opti-MEM for 5 min.
  - iii) After 5 min, they are combined together as a total volume of 200  $\mu$ l and further incubated for 20 min at room temperature.
  - iv) The poly(I:C) packed with Lipofectamine 2000 (A 200  $\mu$ l total at 2.5 mg/ml) is then used to stimulate cells.
4. Incubate at 37°C in a CO<sub>2</sub> incubator for 16 hours.
5. Equilibrate the plate to room temperature for 10 minutes.
6. Add 50  $\mu$ l of luciferase assay reagent (Abeomics, Cat #17-1101; Refer to the reagent datasheet for the detailed luciferase assay protocol) per well.
7. Read the plate in 1-5 minutes to measure luminescence using a microplate luminometer.

## LIMITED USE RESTRICTIONS:

**THIS PRODUCT IS SOLELY FOR IN VITRO RESEARCH USE ONLY. NOT FOR DIAGNOSTIC OR THERAPEUTIC USE.**

**By use of this product, user agrees to be bound by the terms of this limited use statement.**

**This product is solely for Internal Research Purposes and not for Commercial Purposes. Commercial Purposes include, but are not limited to (1) use of the cell line in manufacturing; (2) use of the cell line to provide a service, information or data; (3) use of the cell line for therapeutic, diagnostic or prophylactic purposes; or (4) resale of the cell line whether or not such cell lines are resold for use in research. The buyer cannot sell, give or otherwise transfer this product to a third party.**

**Commercial License Agreement is available for non-research use if applicable. Please contact Abeomics ([info@abeomics.com](mailto:info@abeomics.com)).**

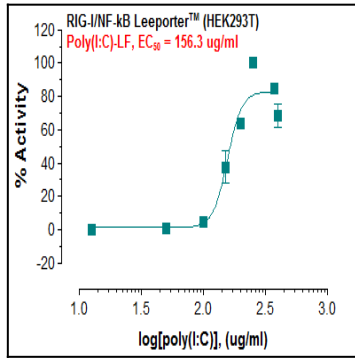


Fig-1: Induction of RIG-I activity by poly(I:C) prepaced with lipofectamine in RIG-I Leepor<sup>™</sup> - HEK293T cells.